

#### **CERTIFICATE OF ANALYSIS**

## HpyF3I (DdeI)

**#ER1882** 2500 u

Lot: Expiry Date:

5'...**C**↓**T N A G**...3' 3'...**G A N T**↑**C**...5'

Concentration: 10 u/µl

Source: E.coli that carries the cloned hpyF3IR

gene from *Helicobacter pylori* RFL3

Supplied with: 1 ml of 10X Buffer Tango

Store at -20°C











In total 2 vials. BSA included: Lot# BSA62-313P



#### RECOMMENDATIONS

**1X Buffer Tango**<sup>™</sup> (for 100% HpyF3I digestion) 33 mM Tris-acetate (pH 7.9), 10 mM magnesium acetate, 66 mM potassium acetate, 0.1 mg/ml BSA.

## **Incubation temperature**

37°C.

#### **Unit Definition**

One unit is defined as the amount of HpyF3I required to digest 1  $\mu$ g of lambda DNA in 1 hour at 37°C in 50  $\mu$ l of recommended reaction buffer.

#### **Dilution**

Dilute with Dilution Buffer (#B19): 10 mM Tris-HCl (pH 7.4 at 25°C), 100 mM KCl, 1 mM EDTA, 1 mM DTT, 0.2 mg/ml BSA and 50% glycerol.

#### **Double Digests**

Tango<sup>™</sup> Buffer is provided to simplify buffer selection for double digests. 98% of Fermentas Restriction Enzymes are active in a 1X or 2X concentration of Tango<sup>™</sup> Buffer. Please refer to the Fermentas Catalog or go to <a href="https://www.fermentas.com/doubledigest">www.fermentas.com/doubledigest</a> to choose the best buffer for your experiments.

### **Storage Buffer**

HpyF3I is supplied in: 10 mM Tris-HCI (pH 7.4 at 25°C), 100 mM KCI, 1 mM EDTA, 1 mM DTT, 0.2 mg/ml BSA and 50% glycerol.

## **Recommended Protocol for Digestion**

• Add:

nuclease-free water	16 µl
10X Buffer Tango <sup>™</sup>	2 µl
DNA (0.5-1 μg/μl)	1 µl
HpyF3I	0.5-2 µl

- Mix gently and spin down for a few seconds.
- Incubate at 37°C for 1-16 hours.

The digestion reaction may be scaled either up or down.

# **Recommended Protocol for Digestion of PCR Products Directly after Amplification**

• Add:

PCR reaction mixture 10  $\mu$ l (~0.1-0.5  $\mu$ g of DNA) nuclease-free water 18  $\mu$ l 10X Buffer Tango 2  $\mu$ l 1-2  $\mu$ l

- Mix gently and spin down for a few seconds.
- Incubate at 37°C for 1-16 hours.

#### **Thermal Inactivation**

HpyF3I is inactivated by incubation at 65°C for 20 min.

#### **ENZYME PROPERTIES**

## **Enzyme Activity in Fermentas REase Buffers, %**

В	G	0	R	Tango <sup>™</sup>	2X Tango <sup>™</sup>
20-50	20-50	20-50	20-50	100	50-100

#### **Methylation Effects on Digestion**

Dam: never overlaps – no effect. Dcm: never overlaps – no effect. CpG: never overlaps – no effect. EcoKI: never overlaps – no effect.

EcoBI: may overlap -effect not determined.

## **Stability during Prolonged Incubation**

A minimum of 0.2 units of the enzyme is required for complete digestion of 1  $\mu$ g of lambda DNA in 16 hours at 37°C.

### **Compatible Ends**

BbvCl, Bpu10l, Bpu1102l, Eco81l.

### **Number of Recognition Sites in DNA**

λ	ФХ174	pBR322	pUC57	pUC18/19	pTZ19R/U	M13mp18/19
104	14	8	6	6	4	29

For **QUALITY CONTROL ASSAY DATA** see back page

#### **QUALITY CONTROL ASSAY DATA**

#### **Overdigestion Assay**

No detectable change in the specific fragmentation pattern is observed after a 160-fold overdigestion with HpyF3I (10 u/µg lambda DNA x 16 hours).

#### **Ligation/Recutting Assay**

After a 50-fold overdigestion (3 u/ $\mu$ g DNA x 17 hours) with HpyF3I more than 95% of the digested DNA fragments can be ligated at a 5'-termini concentration of 1  $\mu$ M. More than 95% of these sites can be recut.

## Labeled Oligonucleotide (LO) Assay

No detectable degradation of single-stranded or doublestranded labeled oligonucleotides occurred during incubation with 10 units of HpyF3I for 4 hours.

#### **Quality authorized by:**



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