

CERTIFICATE OF ANALYSIS

Cfr9I (XmaI)

#ER0172 1500 u

Lot: Expiry Date:

5'...C↓C C G G G...3'
3'...G G G C C↑C...5'

Concentration: 10 u/μl
Source: *E.coli* that carries the cloned *cfr9IR* gene
from *Citrobacter freundii* RFL9
Supplied with: 1 ml of 10X Buffer Cfr9I
1 ml of 10X Buffer Tango™

Store at -20°C



In total 3 vials.

BSA included: Lot# BSA62-313P

RECOMMENDATIONS

1X Buffer Cfr9I (for 100% Cfr9I digestion)

10 mM Tris-HCl (pH 7.2), 5 mM MgCl₂, 200 mM sodium glutamate, 0.1 mg/ml BSA.

Incubation temperature

37°C.

Unit Definition

One unit is defined as the amount of Cfr9I required to digest 1 μg of lambda DNA-HindIII fragments in 1 hour at 37°C in 50 μl of recommended reaction buffer (containing 2μg DNA fragments).

Dilution

Dilute with Dilution Buffer (#B19): 10 mM Tris-HCl (pH 7.4 at 25°C), 100 mM KCl, 1 mM EDTA, 1 mM DTT, 0.2 mg/ml BSA and 50% glycerol.

Double Digests

Tango™ Buffer is provided to simplify buffer selection for double digests. 98% of Fermentas restriction enzymes are active in a 1X or 2X concentration of Tango™ Buffer. Please refer to the Fermentas Catalog or go to www.fermentas.com/doubledigest to choose the best buffer for your experiments.

1X Tango™ Buffer:

33 mM Tris-acetate (pH 7.9 at 37°C), 10 mM magnesium acetate, 66 mM potassium acetate, 0.1 mg/ml BSA.

Storage Buffer

Cfr9I is supplied in: 10 mM Tris-HCl (pH 7.5 at 25°C), 250 mM KCl, 1 mM DTT, 0.1 mM EDTA, 0.2 mg/ml BSA and 50% glycerol.

Recommended Protocol for Digestion

- Add:

nuclease-free water	16 µl
10X Buffer Cfr9I	2 µl
DNA (0.5-1 µg/µl)	1 µl**
Cfr9I	1-2 µl
- Mix gently and spin down for a few seconds.
- Incubate at 37°C for 1-16 hours.

The digestion reaction may be scaled either up or down.

Recommended Protocol for Digestion of PCR Products Directly after Amplification

- Add:

PCR reaction mixture	10 µl** (~0.1-0.5 µg of DNA)
nuclease-free water	18 µl
10X Buffer Cfr9I	2 µl
Cfr9I	1-2 µl
- Mix gently and spin down for a few seconds.
- Incubate at 37°C for 1-16 hours.

** See Note.

Thermal Inactivation

Cfr9I is inactivated by incubation at 65°C for 20 min.

ENZYME PROPERTIES

Enzyme Activity in Fermentas REase Buffers, %

Cfr9I	B	G	O	R	Tango™	2X Tango™
100	0-20	0-20	0-20	0-20	20-50	0-20

Methylation Effects on Digestion

Dam: never overlaps – no effect.
Dcm: never overlaps – no effect.
CpG: completely overlaps – cleavage impaired.
EcoKI: never overlaps – no effect.
EcoBI: never overlaps – no effect.

Stability during Prolonged Incubation

A minimum of 0.2 units of the enzyme is required for complete digestion of 1 µg of lambda DNA in 16 hours at 37°C.

Digestion of Agarose-embedded DNA

A minimum of 5 units of the enzyme is required for complete digestion of 1 µg of agarose-embedded lambda DNA in 16 hours.

Compatible Ends

BshTI, BsaWI, Cfr10I, Eco88I, Kpn2I, NgoMIV, SgrAI.

Number of Recognition Sites in DNA

λ	ΦX174	pBR322	pUC57	pUC18/19	pTZ19R/U	M13mp18/19
3	0	0	1	1	1	1

Note

To achieve complete digestion of substrate with Cfr9I, the concentration of DNA in reaction buffer should not be less than 50 µg/ml.

For **QUALITY CONTROL ASSAY DATA** see back page

QUALITY CONTROL ASSAY DATA

Overdigestion Assay

No detectable change in the specific fragmentation pattern is observed after a 160-fold overdigestion with Cfr 9I (10 u/μg lambda DNA x 16 hours).

Ligation/Recutting Assay

After a 50-fold overdigestion (3 u/μg DNA x 17 hours) with Cfr 9I, more than 95% of the digested DNA fragments can be ligated at a 5'-termini concentration of 0.03 μM. More than 95% of these sites can be recut.

Labeled Oligonucleotide (LO) Assay

No detectable degradation of single-stranded or double-stranded labeled oligonucleotides occurred during incubation with 10 units of Cfr 9I for 4 hours.

Quality authorized by:



Jurgita Zilinskiene

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